

# INSTRUCTION MANUAL

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## **SERVA***Light Vega*

**Western Blot Chemiluminescence HRP Substrate Kit**

**(Cat. No. 42588)**



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# 1. **SERVALight Vega**

## 1.1. **General information**

**SERVALight Vega** is an extremely sensitive enhanced chemiluminescent substrate for detecting horseradish peroxidase (HRP) conjugates on immunoblots. The extremely intense signal output enables detection of **low picogram** ( $10^{-12}$ ) amounts of antigen. The sensitivity, intensity and duration of the signal allow easy detection of HRP using imaging equipment.

Advantages:

- Compatible with other chemiluminescence detection protocols, e.g. ECL Amesham, ECL Pierce.
- Detection with films or digitally by using documentation systems suitable for chemiluminescence, e.g. ProXima C16Phi+, Isogen
- Very short exposure times and/or highly diluted antibody solution because of extremely intense light output

**SERVALight Vega** is intended for research use only, and shall not be used in any clinical procedures, or for diagnostic purposes.

SERVALight is produced by Cyanagen Srl. Cyanagen Srl is subject of US and EU patent application number US7803573; EP1962095; US7855287; EP1950207, together with other equivalent granted patents and patent applications in other countries.

## 1.2. **Kit components**

| <b>Cat. No.</b> | <b>SERVALight Vega Luminol Solution</b> | <b>SERVALight Vega Peroxide Solution</b> |
|-----------------|---|--|
| 42588.01        | 25 ml                                   | 25 ml                                    |
| 42588.02        | 125 ml                                  | 125 ml                                   |
| 42588.03        | 250 ml                                  | 250 ml                                   |

## 1.3. **Storage conditions**

Upon arrival store the kit at 2 – 8 °C.

If stored at the recommended temperature at least usable until: see expiry date on package.

## 2. Detection procedure

### 2.1. Important before starting

**Safety information:**

*For safety reasons always wear suitable protective gloves and clothing, when you work in the lab.*

It is essential to optimize all components of the system including sample amount, primary and secondary antibody concentration, and the choice of membrane as well as blocking reagents. Therefore, the recommended antibody dilutions has to be critically adapted. Due to this high sensitivity, the substrate requires fewer amounts of sample, primary and secondary antibodies than most commercially available substrates.

- The antibody solutions required will be much more dilute than those used with colorimetric HRP systems. To optimize the appropriate concentrations, perform a systematic dot blot analysis.
- No blocking reagent is optimal for all systems Therefore, it is important to determine the appropriate blocking buffer for each Western blot system. Determining the proper blocking buffer can increase sensitivity and prevent non-specific signal caused by cross-reactivity between the antibody and the blocking reagent. Furthermore, when switching from one substrate to another, a diminished signal or increased background can result if the blocking buffer is not optimal for the new system.
- Use a sufficient volume of wash buffer, blocking buffer, antibody solution and the substrate working solution to cover blot and ensure that it never becomes dry. Large blocking and wash buffer volumes result in reduced non-specific signal.
- Add Tween<sup>®</sup>-20 (final concentration of 0.05%) to the blocking buffer and when preparing all antibody dilutions to reduce non-specific signal. Use only high-quality products guaranteed to be low in peroxides and other contaminants.
- Do not use milk as a blocking reagent when using avidin/biotin systems because milk contains variable amounts of endogenous biotin.
- For optimal results, use a shaking platform during incubation steps.
- Do not use sodium azide as a preservative for buffers. Sodium azide is an inhibitor of HRP and could interfere with this system.

- Do not handle membrane with bare hands. Always wear gloves or use clean forceps. All equipment must be clean and free of foreign material. Metallic devices, e.g. scissors must have no visible signs of rust. Rust may cause speckling and/or high background
- **SERVALight Vega** substrate working solution is stable for 5 day at room temperature. Exposure to the sun or any other intense light can harm the working solution. For best results keep the working solution in an amber bottle and do not expose to any intense light. Short-term exposure to typical laboratory lighting will not harm the working solution.

## 2.2. Required, not supplied materials and solutions

- **Western-Blot-Membran:** Use any suitable protocol to separate proteins by electrophoresis and transfer them to a nitrocellulose membrane.
- **Dilution buffer:**  
TBS (Tris Buffered Saline) or PBS (Phosphate Buffered Salin)
- **Blocking buffer:**  
Suitable blocking in TBS oder PBS
- **Wash buffer (TBS-T or PBS-T):**  
5 ml 10 % (v/v) Tween<sup>®</sup>-20  
ad 1 l TBS or PBS
- **Blocking reagent:**  
0.5 ml 10 % (v/v) Tween<sup>®</sup>-20  
ad 100 ml blocking buffer (with the same base components as the dilution buffer)
- **Primary antibody (1. Ab, target protein specific antibody):**
  - Stock solution in dilution buffer: 1 mg/ml
  - Working solution: 1: 100 – 1: 5,000 dilution in blocking reagent.

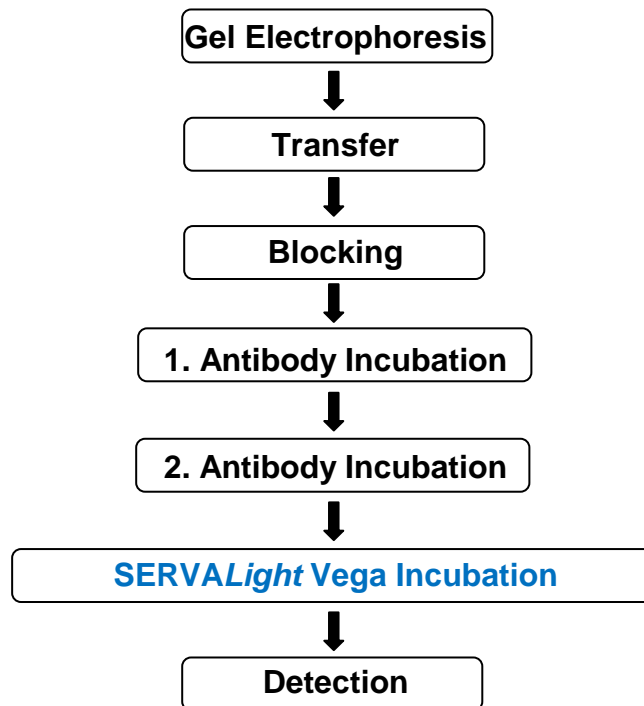
The necessary dilution to use depends on the specific primary antibody and the amount of antigen on the membrane and will require optimization for each experimental system.

- **Secondary antibody HRP-labelled (2. Ab, specific for primary antibody):**
  - Stock solution in blocking reagent: 1 mg/ml
  - Working solution: 1: 1,000 – 1: 15,000 dilution in blocking reagent.

The optimal dilution varies depending on the HRP conjugate and the amount of antigen on the membrane.

- Rotary platform shaker: For agitation of membrane during incubations.
- Film cassette, developing and fixing reagents: For processing auto-radiographic film.

### 2.3. Overview of the Western Blot procedure



### 2.4. Protocol

- (1) Remove blot from the transfer apparatus and block non-specific sites with blocking reagent for 30 - 60 min at room temperature (RT) with shaking. For best results, block for 1 h at RT or overnight at 2 – 8 °C.
- (2) Remove the blocking reagent and add the appropriate primary antibody dilution. Incubate blot for 1 – 2 h with shaking or overnight at 2 - 8 °C.
- (3) Wash membrane by suspending it in wash buffer and agitating for 5 min. Replace wash buffer at least 4 - 6 times. Increasing the wash buffer volume and/or the number of washes may help reduce background. Briefly rinsing membrane in wash buffer before incubation will increase wash efficiency.
- (4) Add the appropriate HRP conjugate dilution. Incubate blot for 30 – 60 min at RT with shaking. Increasing incubation time will lead to higher background.

- (5) Repeat Step 3 to remove unbound HRP conjugate.  
Note: Membrane must be thoroughly washed after incubation with the HRP conjugate.
- (6) Allow the detection solutions to equilibrate to 20 - 25°C temperature before opening.
- (7) Prepare the substrate working solution by mixing equal parts of the peroxide solution and the luminol solution. Use 0.1 ml of working solution per cm<sup>2</sup> of membrane.
- (8) The working solution is stable for 5 days at 2 - 8 °C.
- (9) Incubate blot with the substrate working solution for 1 min.
- (10) Place the protected blot (e.g. air-bubble-free in saran wrap) in a film cassette with the protein side facing up. Turn off all lights except those appropriate for film exposure (e.g. a red light). Film must remain dry during exposure. For optimal results, perform the following precautions: make sure, excess substrate is removed from the blot and the membrane protector. Use gloves during the entire film-handling process. Never place a blot on developed film. There may be chemicals on the film that will reduce signal.
- (11) Carefully place film on top of the protected blot.
- (12) A recommended exposure time to start with is 30 s, however, exposure time can be varied, e.g. 15 s, 1 min, 5 min, to achieve optimal results. Enhanced or pre-flashed auto-radiographic film is not necessary.  
**Caution:** Light emission is intense and any movement between the film and the blot can cause artefacts on the film.
- (13) Develop film using appropriate developing solution and fixative.
- (14) Blot may be stripped and re-probed if necessary.

### 3. Trouble shooting

| <b>Problem</b>                    | <b>Possible cause</b>   | <b>Countermeasure</b>                               |
|-----------------------------------|---|---|
| <b>Weak or no signal</b>          | Too much HRP in the system depleted the substrate and caused the signal to fade quickly | Higher dilution of HRP conjugate                    |
|                                   | Insufficient quantities of antigen or antibody  | Increase amount of antigen or antibody              |
|                                   | Inefficient protein transfer  | Optimize transfer                                   |
|                                   | Reduction of HRP or substrate activity  | Use fresh reagents                                  |
| <b>Blot glows in the dark</b>     | Too much HRP in the system depleted the substrate                                       | Higher dilution of HRP conjugate                    |
| <b>Membrane has brown bands</b>   |   |   |
| <b>Reverse image on film</b>      |   |   |
| <b>High background</b>            | Too much HRP in the system depleted the substrate                                       | Higher dilution of HRP conjugate                    |
|                                   | Inadequate blocking or inappropriate blocking reagent                                   | Optimize blocking conditions and reagent            |
|                                   | Film over exposed   | Reduce exposure time                                |
|                                   | Too much antigen or antibody  | Optimize amount of antigen or antibody              |
|                                   | Inadequate washing  | Increase length, number and volume of washing steps |
| <b>Speckled background</b>        | Formation of aggregates in HRP conjugate  | Filter through 0.2 µm filter                        |
| <b>Spots within protein bands</b> | Inefficient protein transfer  | Optimize transfer                                   |
|                                   | Unevenly hydrated membrane  | Hydrate the membrane properly                       |
|                                   | Bubbles between film and membrane   | Remove all air bubbles before exposure to film      |
|                                   | Too much antigen or antibody  | Optimize amount of antigen or antibody              |
|                                   | Inadequate washing  | Increase length, number and volume of washing steps |
| <b>Non-specific bands</b>         | Short signal duration and high background (too much HRP)                                | Higher dilution of HRP conjugate                    |
|                                   | Signal duration and background OK (too much primary antibody)                           | Higher dilution of primary antibody                 |
|                                   | SDS present in the immunoassay  | Do not use SDS                                      |



## 4. Ordering information

| <b>Membrans</b>   | <b>Cat. No.</b> |
|---|-----------------|
| Immobilon™ (PVDF), 26.5 cm x 3.75 m, Pore size: 0.2 µm (1 roll) | 42574           |
| Fluorobind (PVDF), 10 x 10 cm, Pore size: 0.2 µm (20 sheets)    | 42573           |
| Fluorobind (PVDF), 25 cm x 3 m, Pore size: 0.2 µm (1 roll)      | 42571           |
| <b>Protein Standard</b>   |                 |
| SERVA Western Blot Protein Standard                             | 39256           |
| <b>Reagents</b>   |                 |
| Tween® 20   | 37470           |
| <b>Detection reagents/-kits</b>                                 |                 |
| Chemiluminescence Reagent for Horseradish Peroxidase            | 42582           |
| SERVALight Polaris CL HRP WB Substrate Kit                      | 42584           |
| SERVALight Eos CL HRP WB Substrate Kit                          | 42585           |
| SERVALight EosUltra CL HRP WB Substrate Kit                     | 42586           |
| SERVALight Helios CL HRP WB Substrate Kit                       | 42587           |

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